SUPPLEMENTARY MATERIAL

We list the methods to measure each individual marker below:

Inflammation

- *Adiponectin (μg/ml):* we measured total plasma adiponectin with a commercial enzyme-linked immunosorbent assays (ELISA) (R&D Systems). The inter-assay coefficients of variation for the assay was < 7%.
- *C-reactive protein (CRP) (mg/l):* we measured plasma CRP with high-sensitivity immunonephelometry (Prospec, Dade Behring Milton Keynes, UK) following the manufacturer's reagents and standards. Intra- and inter-assay coefficients of variation were 4.7% and 8.3%, respectively.
- *Intercellular adhesion molecule-1 (ICAM-1) (ng/ml):* we measured plasma ICAM-1 with a commercially available ELISA (R&D Systems, Abingdon, UK). The inter-assay coefficient of variation was <7%.
- *Interleukin-6 (IL-6) (pg/ml):* we assayed serum IL-6 with a high sensitivity ELISA (R & D Systems, Abingdon, UK). Intra- and inter-assay coefficients of variation were 7.5% and 8.9%, respectively.
- *Tumour necrosis factor-alpha (TNF-α): (pg/ml):* we assayed serum TNF- α with a high-sensitivity ELISA (R&D Systems, Abingdon, UK). Intra-assay and inter assay coefficients of variation were 8.4% and 12.5%, respectively.
- *Interleukin-10 (IL-10) (pg/ml):* we measured serum IL10 with an ELISA assay (R&D Systems, Abingdon, UK). The inter-assay coefficient of variation was 4.5%.
- *Matrix-metalloproteinase-9 (MMP-9) (ng/ml):* we measured serum MMP-9 with a commercially available sandwich ELISA (R&D Systems, Abingdon, UK). Intra-assay and inter assay coefficients variation were 4.4% and 10.4%, respectively.
- *von Willebrand factor (vWF) (IU/dL):* we measured serum vWF antigen with an ELISA using rabbit antihuman polyclonal antibodies obtained from DAKO (High Wycombe, UK). Intra and inter assay coefficient of variation were 3.3% and 4.2%, respectively.

Clotting

- *D-dimer (ng/ml):* we measured plasma levels of fibrin D-dimer with a commercially available ELISA from Biopool AB, Umea Sweden. The intra and inter assay coefficients of variation were 4.7 and 5.2%, respectively.
- *Fibrinogen (g/l):* we measured fibrinogen in plasma by imunonephelometry (Prospec, Dade Behring Milton Keynes, UK) using the manufacturer's reagents and standards. Intra- and inter-assay coefficients of variation were 7.5 and 8.9%, respectively.

Thrombolysis

- *Tissue plasminogen activator (tPA) (ng/mL):* we measured plasma levels of tissue plasminogen activator (t-PA) antigen with commercially available ELISAs from Biopool AB, Umea Sweden. The intra and inter assay coefficients of variation were 6.6% and 6.5%, respectively.

Cardiac Strain

- *N-terminal-pro-brain-natriuretic-peptide (NT pro-BNP) (pg/ml):* we measured serum levels of NT pro-BNP using the Elecsys 2010 electrochemiluminescence analyser (Roche Diagnostics, Burgess Hill, UK) calibrated using the manufacturer's reagents. Manufacturer's controls were used with limits of acceptability defined by the manufacturer. Low control coefficient of variation was 6.7% and high control coefficient of variation was 4.9%.
- *Troponin T (ng/ml):* we determined serum troponin T using the Elecsys 2010 electrochemiluminescence analyser (Roche Diagnostics, Burgess Hill, UK) calibrated using the manufacturer's reagents. Manufacturer's control was used with limits of acceptability defined by the manufacturer. The abnormal high (detectable) control coefficient of variation was 2.3%.

Cerebral damage

- *Tau (pg/ml):* we measured tau in serum with a sandwich ELISA using the Innotest htau antigen (Innogenetics). The coefficient of variation at 479pg/ml was 5.8%.
- *S100 B (pg/ml):* we measured S100 B in serum. 96-well microtiter plates were coated with 200 μL of 0.05 M carbonate buffer containing monoclonal anti-S100 B (Affiniti Research Products, Exeter, UK). The plates were washed with 0.67 M barbitone buffer containing 5 mM calcium lactate, 0.1% bovine serum albumin, and 0.05% Tween and then were blocked with 2% bovine serum albumin and washed again. Two-hundred microliters of diluted

serum (1:1) in 0.67 M barbitone buffer containing 5 mM calcium lactate was added in duplicate. After incubation and washing, horseradish-peroxidase-conjugated polyclonal anti-S100 B (Dako, Copenhagen, Denmark) was used as a detecting antibody. The o-phenylenediamine color reaction was stopped with 1 M hydrochloric acid, and the absorbances were read at 492 and 405 nm. The antigen concentration was calculated from an internal standard curve ranging from 0 to 250 pg/mL. The coefficient of variation at a concentration of 263pg/ml was 11%.